

Screening for trisomies by cell-free DNA testing of maternal blood: consequences of a failed result

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KEYWORDS: cell-free DNA; fetal fraction; first-trimester screening; non-invasive prenatal testing; trisomy 21

ABSTRACT

Objectives First, to report the distribution of the fetal fraction of cell-free (cf) DNA and the rate of a failed cfDNA test result in trisomies 21, 18 and 13, by comparison with pregnancies unaffected by these trisomies, second, to examine the possible effects of maternal and fetal characteristics on the fetal fraction, and third, to consider the options for further management of pregnancies with a failed result.

Methods This was a cohort study of 10 698 singleton pregnancies undergoing screening for fetal trisomies 21, 18 and 13 by cfDNA testing at 10–14 weeks' gestation. There were 160 cases of trisomy 21, 50 of trisomy 18, 16 of trisomy 13 and 10 472 were unaffected by these trisomies. Multivariate regression analysis was used to determine significant predictors of fetal fraction and a failed cfDNA test result amongst maternal and fetal characteristics.

Results Fetal fraction decreased with increasing body mass index and maternal age, was lower in women of South Asian racial origin than in Caucasians and in assisted compared to natural conceptions. It increased with fetal crown–rump length and higher levels of serum pregnancy-associated plasma protein-A and free β -human chorionic gonadotropin. The median fetal fraction was 11.0% (interquartile range (IQR), 8.3–14.4%) in the unaffected group, 10.7% (IQR, 7.8–14.3%) in trisomy 21, 8.6% (IQR, 5.0–10.2%) in trisomy 18 and 7.0% (IQR, 6.0–9.4%) in trisomy 13. There was a failed result from cfDNA testing after first sampling in 2.9% of the unaffected group, 1.9% of trisomy 21, 8.0% of trisomy 18 and 6.3% of trisomy 13. In the cases with a failed result, 7% of women had invasive testing, mainly because of high risk from the combined test and/or presence of sonographic features suggestive of trisomies 18 and 13. All cases of trisomies were detected prenatally.

Conclusions In cases of a failed cfDNA test, the rate of trisomies 18 and 13, but not trisomy 21, is higher than in those with a successful test. In the management of such cases, the decision in favor of invasive testing should depend on the risk of prior screening and the results of detailed ultrasound examination. Copyright © 2016 ISUOG. Published by John Wiley & Sons Ltd.

INTRODUCTION

Cell-free (cf) DNA analysis of maternal blood provides effective screening for fetal trisomies 21, 18 and 13 with reported detection rates (DR) of 99%, 96% and 91%, respectively, at an overall false-positive rate (FPR) of 0.35%¹. However, the test fails to provide a result in up to 8% of cases and the most common reason for such failure is low fetal fraction². There are some limited data indicating that, in the pregnancies with failed results, fetal chromosomal abnormalities are over-represented² and this has led to a recommendation by the American College of Obstetricians and Gynecologists (ACOG) that, in cases of a failed result, women should be offered diagnostic testing³.

The objectives of this cohort study of 10 698 singleton pregnancies undergoing screening for fetal trisomies 21, 18 and 13 by cfDNA testing at 10–14 weeks' gestation were first, to report the distribution of fetal fraction of cfDNA and the rate of a failed result in each of the trisomies and compare with pregnancies unaffected by these trisomies, second, to examine the possible effects of maternal and fetal characteristics on the fetal fraction and third, to consider the options for the further management of pregnancies with a failed cfDNA test result.

METHODS

The data for this study were derived from first, cfDNA testing as an option following first-trimester combined

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Accepted: 31 December 2015

testing in women with singleton pregnancies attending for routine care at 11+0 to 13+6 weeks' gestation in one of two National Health Service (NHS) hospitals in England⁴ and second, cfDNA testing as part of routine screening in women with singleton pregnancies at 10+0 to 13+6 weeks attending the Fetal Medicine Centre in London, which is a private clinic⁵. The patients were examined between October 2012 and August 2015.

We recorded maternal characteristics and medical history, including maternal age, racial origin (Caucasian, African, South Asian, East Asian and mixed), method of conception (natural/assisted conception requiring the use of ovulation drugs/*in-vitro* fertilization), cigarette smoking during pregnancy (yes/no) and parity (parous/nulliparous if no previous pregnancy at or after 24 weeks' gestation). We also measured maternal weight and height. In all cases, free beta-human chorionic gonadotropin (β -hCG) and pregnancy-associated plasma protein-A (PAPP-A) were measured within 10 min of blood collection at 10+0 to 13+6 weeks (DELFLIA Xpress system, PerkinElmer Life and Analytical Sciences, Waltham, USA, or Kryptor, Thermo Scientific, Berlin, Germany). An ultrasound scan was carried out at 11+0 to 13+6 weeks to determine gestational age from the measurement of the fetal crown-rump length (CRL)⁶, diagnose any major fetal abnormalities and measure fetal nuchal translucency (NT) thickness. The measured NT was expressed as a difference from the expected normal mean for gestation (delta value)⁷. Similarly, the measured free β -hCG and PAPP-A were converted into multiples of the median (MoM) for gestational age, adjusted for maternal weight, racial origin, smoking status, method of conception, parity and machine used for the assays⁸. Biophysical and biochemical markers were combined to estimate the patient-specific risk for trisomies 21, 18 and 13.

Women provided written informed consent and maternal blood (20 mL) was sent via courier to the USA for cfDNA testing (HarmonyTM Prenatal Test, Ariosa Diagnostics, Inc., San Jose, CA, USA)⁹⁻¹³. Chromosome-selective sequencing, referred to as digital analysis of selected regions (DANSR), and fetal-fraction optimized risk of trisomy evaluation (FORTETM) were used to assay non-polymorphic and polymorphic loci, where fetal alleles differ from maternal alleles, enabling simultaneous determination of chromosome proportion and fetal fraction. The results from cfDNA testing were presented as risk scores for trisomy 21, 18 and 13 which in most cases were either > 99% or < 1:10 000. In cases for which the cfDNA test did not provide results the parents were offered repeat testing or to rely on the results of the combined test in deciding whether to have an invasive test or not. In cases with a high-risk result from the cfDNA test, the parents were advised to consider having invasive fetal karyotyping before deciding on the further management of their pregnancy.

Patient characteristics and results of the investigations were recorded in a fetal database. Results

from invasive testing, obtained from laboratories, and pregnancy outcome, obtained from obstetricians, general practitioners or the patients, were recorded in the same database. The outcomes were divided into first, trisomy 21, 18 or 13 if the karyotype of chorionic villi, amniotic fluid or neonatal blood demonstrated the relevant trisomy, second, no trisomy 21, 18 or 13 if the karyotype of chorionic villi, amniotic fluid or neonatal blood was normal or the neonate was phenotypically normal, third, unknown karyotype because the pregnancy resulted in miscarriage or stillbirth and no karyotyping of fetal tissue was carried out, and fourth, outcome unknown because the pregnancy was lost to follow up.

Statistical analysis

Descriptive data are presented as median and interquartile range (IQR) for continuous variables and as numbers and percentages for categorical variables. The measured fetal fraction was \log_{10} transformed to make the distribution Gaussian, which was assessed using histograms and probability plots. Univariate and multivariate regression analyses were used to determine which of the factors amongst maternal age, body mass index, racial origin, smoking status, method of conception, fetal CRL, serum PAPP-A and free β -hCG, fetal NT and fetal karyotype were significant predictors of \log_{10} fetal fraction. In each trisomic and unaffected pregnancy the \log_{10} fetal fraction was expressed as a MoM after adjusting for the maternal variables found to be significant in the multivariate regression analysis. Logistic regression analysis was undertaken to examine the maternal and pregnancy characteristics providing significant contribution to prediction of a failed cfDNA test result.

The statistical software package SPSS 21.0 (SPSS Inc., Chicago, IL, USA) was used for data analyses.

RESULTS

Characteristics of study population

A total of 10 963 women had cfDNA testing and combined screening for trisomies, but 265 (2.4%) of these were excluded from further analysis either because the pregnancy ended in termination, miscarriage or stillbirth with no known karyotype ($n = 155$), was lost to follow up ($n = 85$) or had a chromosomal abnormality other than trisomy 21, 18 or 13 ($n = 25$).

Maternal and pregnancy characteristics in the 10 698 cases with known outcome are summarized in Table 1. There were 160 cases of trisomy 21, 50 of trisomy 18, 16 of trisomy 13 and 10 472 were unaffected by these trisomies. Results from cfDNA testing were provided after first sampling for 97.0% (10 382/10 698) of cases, including 97.1% in the unaffected group, 98.1% in trisomy 21, 92.0% in trisomy 18 and 93.7% in trisomy 13. The reasons for failure to provide a result were low fetal fraction in 219 (69.3%) cases and laboratory processing problems in 97 (30.7%) cases.

Table 1 Maternal and pregnancy characteristics of the study population of 10 698 women with singleton pregnancy undergoing cell-free DNA testing for trisomies 21, 18 and 13

Characteristic	Value (n = 10 698)
Maternal age (years)	36.3 (33.2–39.3)
Maternal weight (kg)	64.0 (57.9–73.0)
Maternal height (cm)	165 (161–170)
Racial origin	
Caucasian	8751 (81.8)
African	698 (6.5)
South Asian	663 (6.2)
East Asian	386 (3.6)
Mixed	200 (1.9)
Nulliparous	4760 (44.5)
Cigarette smoker	263 (2.5)
Mode of conception	
Natural	9515 (88.9)
Assisted	1183 (11.1)
Crown–rump length (mm)	53.7 (38.5–65.7)

Data are given as median (interquartile range) or *n* (%).

Factors affecting fetal fraction

At first sampling, the median fetal fraction was 11.0% (IQR, 8.3–14.4%) in the unaffected group, 10.7% (IQR, 7.8–14.3%) in trisomy 21, 8.6% (IQR, 5.0–10.2%) in trisomy 18 and 7.0% (IQR, 6.0–9.4%) in trisomy 13; in these calculations, it was assumed that in the cases with a failed result because of low fetal fraction, the fetal fraction was 3%.

Log₁₀ fetal fraction from first sampling had a Gaussian distribution (Figure 1). Univariate regression analysis demonstrated that significant independent prediction of log₁₀ fetal fraction was provided by maternal age, body mass index, African, South Asian and East Asian racial origins, assisted conception, MoM values of PAPP-A and free β-hCG and trisomies 18 and 13 (Table 2). In the multivariate regression analysis, significant contribution was provided by maternal age, body mass index, South Asian racial origin, assisted conception, fetal CRL and MoM values of PAPP-A and free β-hCG, but not trisomies 18 or 13 (adjusted $R^2 = 0.251$; $P < 0.0001$). If in the multivariate regression analysis we excluded PAPP-A and free β-hCG MoM, significant contribution to log₁₀ fetal fraction was provided by maternal age, body mass index, South Asian racial origin, assisted conception, fetal CRL and trisomies 18 and 13 (adjusted $R^2 = 0.174$; $P < 0.0001$).

We used the coefficients of the maternal characteristics with significant contribution to log₁₀ fetal fraction in the multivariate regression analysis to derive a model for calculation of MoMs. The median fetal fraction MoM in unaffected pregnancies was 1.03 (IQR, 0.79–1.32). Compared to unaffected pregnancies, the fetal fraction MoM was not significantly different in trisomy 21 (0.99 (IQR, 0.77–1.29), $P = 0.527$), but it was lower in those with trisomy 18 (0.80 (IQR, 0.49–1.05), $P < 0.0001$) or trisomy 13 (0.71 (IQR, 0.54–0.90), $P < 0.0001$) (Figure 2). If in the estimation of fetal fraction MoM we included the coefficients for PAPP-A and free β-hCG

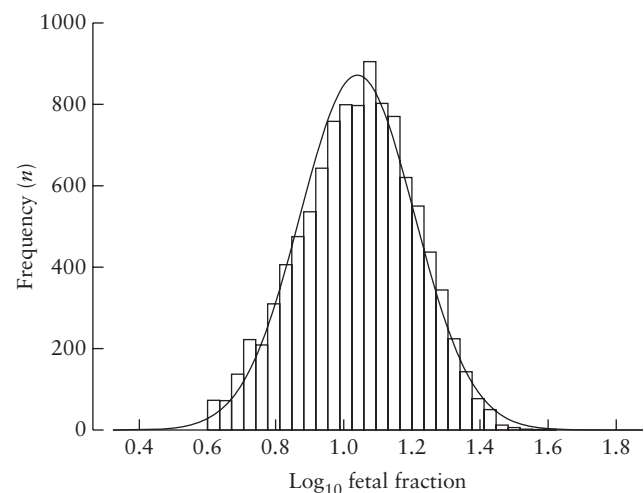


Figure 1 Frequency distribution of log₁₀ fetal fraction of cell-free DNA in maternal blood from first sampling in 10 698 pregnant women.

MoM, there were no significant differences between the unaffected pregnancies (1.03 (IQR, 0.81–1.30)) and those with trisomy 21 (1.05 (IQR, 0.76–1.29), $P = 0.894$), trisomy 18 (1.10 (IQR, 0.77–1.52), $P = 0.198$) or trisomy 13 (0.81 (IQR, 0.66–1.14), $P = 0.051$).

Multivariate logistic regression analysis demonstrated that the risk of test failure increased with increasing maternal age and body mass index, decreased with increasing PAPP-A and free β-hCG MoM and was higher in women of South Asian than Caucasian racial origin and in pregnancies achieved by assisted conception than those achieved naturally (adjusted $R^2 = 0.212$; $P < 0.0001$).

Management of pregnancies with cfDNA test failure

There was a failed cfDNA result after first sampling in 2.9% (308/10 472) of cases in the unaffected group, in 1.9% (3/160) with trisomy 21, 8.0% (4/50) with trisomy 18 and 6.3% (1/16) with trisomy 13 (Table 3).

In the 308 unaffected cases with a failed cfDNA result, 234 (76.0%) chose repeat cfDNA testing, eight (2.6%) had invasive testing and 66 (21.4%) opted for no further investigations. Repeat cfDNA testing provided a result in 147 (62.8%) of the 234 cases; seven (8.0%) of the 87 with a failed second cfDNA test had invasive testing and 80 (92.0%) opted for no further investigations. In total, 15 (4.9%) of the 308 women with a failed cfDNA result ended up having an invasive test and in 13 (86.7%) of the 15 cases in this group the estimated risk for trisomies from the combined test was ≥ 1 in 100. In contrast, in the 146 cases with no result from first or repeat cfDNA testing and who decided against invasive testing, only 14 (9.6%) had an estimated risk for trisomies from the combined test of ≥ 1 in 100.

In two of the three cases of trisomy 21 with a failed cfDNA result, invasive testing was carried out because the estimated risk from the combined test was ≥ 1 in 3;

Table 2 Univariate and multivariate regression analysis demonstrating factors from maternal and pregnancy characteristics that contribute significantly to prediction of log₁₀-transformed fetal fraction

Independent variable	Univariate		Multivariate	
	Regression coefficient	P	Regression coefficient	P
Intercept			1.451 (1.419 to 1.483)	< 0.0001
Age in years	-0.003 (-0.004 to -0.002)	< 0.0001	-0.002 (-0.003 to -0.001)	< 0.0001
BMI in kg/m ²	-0.016 (-0.016 to -0.015)	< 0.0001	-0.016 (-0.017 to -0.015)	< 0.0001
Racial origin				
Caucasian (reference)	0.000			
African	-0.055 (-0.070 to -0.039)	< 0.0001		
South Asian	-0.021 (-0.036 to -0.005)	0.011	-0.019 (-0.032 to -0.005)	0.008
East Asian	0.029 (0.009 to 0.050)	0.005		
Mixed	0.022 (-0.006 to 0.051)	0.123		
Cigarette smoking	0.013 (-0.011 to 0.038)	0.293		
Assisted conception	-0.089 (-0.101 to -0.077)	< 0.0001	-0.086 (-0.097 to -0.075)	< 0.0001
Fetal CRL in mm	1.1E ⁻⁰⁴ (1.1E ⁻⁰⁴ to 3.2E ⁻⁰⁴)	0.412	0.001 (4.7E ⁻⁰⁴ to 0.001)	< 0.0001
Log ₁₀ PAPP-A MoM	0.166 (0.152 to 0.180)	< 0.0001	0.133 (0.119 to 0.146)	< 0.0001
Log ₁₀ free β-hCG MoM	0.171 (0.158 to 0.184)	< 0.0001	0.140 (0.128 to 0.152)	< 0.0001
Delta NT	-0.003 (-0.009 to 0.002)	0.251		
Fetal karyotype				
Trisomy 21	0.003 (-0.028 to 0.035)	0.837		
Trisomy 18	-0.142 (-0.198 to -0.085)	< 0.0001		
Trisomy 13	-0.161 (-0.260 to -0.063)	< 0.0001		

Values in parentheses are 95% CIs. β-hCG, beta-human chorionic gonadotropin; CRL, crown-rump length; MoM, multiples of the median; NT, nuchal translucency; PAPP-A, pregnancy-associated plasma protein-A.

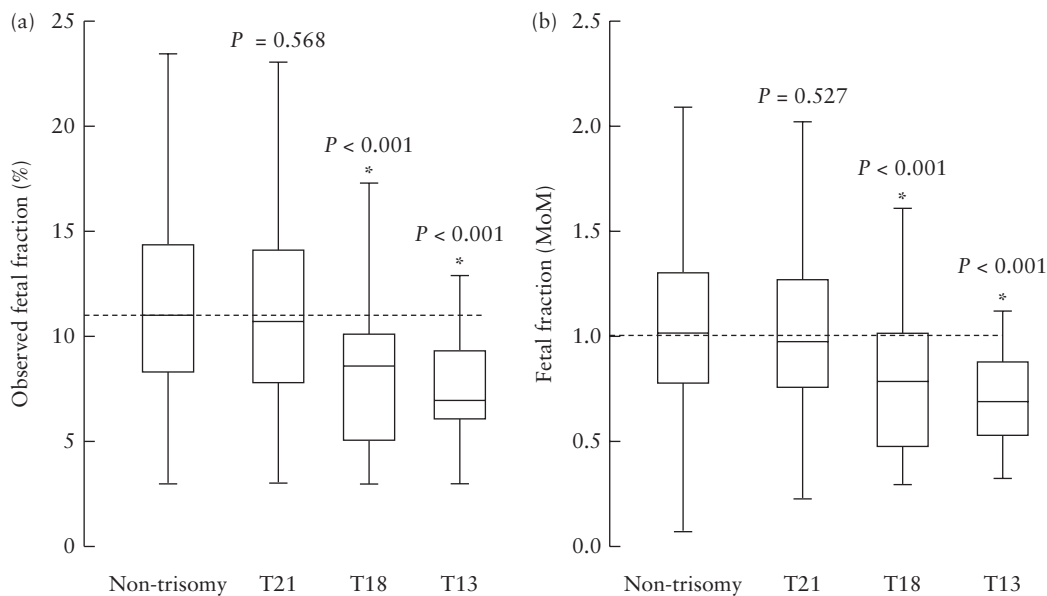


Figure 2 Box-and-whisker plot of fetal fraction (a) and fetal fraction multiples of the median (MoM) (b) in normal pregnancies and those with trisomies 21 (T21), 18 (T18) or 13 (T13). Boxes represent median and interquartile range, and whiskers represent range. MoMs were calculated from coefficients in Table 2. Comparison with normal pregnancy: **P* < 0.05.

in the third case, with an estimated risk of 1 in 13, the cfDNA test was repeated and this gave a result. In the four cases of trisomy 18 with a failed cfDNA result, invasive testing was carried out because in all cases the estimated risk from the combined test was ≥ 1 in 5, serum PAPP-A and free β-hCG was ≤ 0.3 MoM and there were sonographic features suggestive of this trisomy, including clenched hands, cardiac defect and/or exomphalos. In the case of trisomy 13 with a failed cfDNA test result, invasive testing was carried out because the estimated risk from

the combined test was 1 in 2, the fetal NT was 4.1 mm and the fetal heart rate was 200 bpm.

DISCUSSION

Principal findings of the study

In this study, selective sequencing was used for cfDNA analysis of maternal blood in screening for fetal trisomies 21, 18 and 13 at 10–14 weeks' gestation. The median fetal

Table 3 Results from cell-free DNA test and further management of pregnancies according to trisomic status of the fetus

	No trisomy	Trisomy 21	Trisomy 18	Trisomy 13
Number of cases	10 472	160	50	16
Median fetal fraction				
Percentage	11.0 (8.3–14.4)	10.7 (7.8–14.3)	8.6 (5.0–10.2)	7.0 (6.0–9.4)
MoM	1.03 (0.79–1.32)	0.99 (0.77–1.29)	0.80 (0.49–1.05)	0.71 (0.54–0.90)
Failed result	308/10 472 (2.9)	3/160 (1.9)	4/50 (8.0)	1/16 (6.3)
Low fetal fraction	214/308 (69.5)	2/3 (66.7)	2/4 (50.0)	1/1 (100)
Laboratory processing	94/308 (30.5)	1/3 (33.3)	2/4 (50.0)	
Response to failed result				
Invasive testing	8/308 (2.6)	2/3 (66.7)	4/4 (100)	1/1 (100)
No further testing	66/308 (21.4)			
Repeat testing	234/308 (76.0)	1/3 (33.3)		
Result	147/234 (62.8)	1/1 (100)		
Failed result	87/234 (37.2)			
Invasive testing	7/87 (8.0)			
No further testing	80/87 (92.0)			

Values are given as *n*, median (interquartile range) or *n/N* (%). MoM, multiples of the median.

fraction was 11.0% and this decreased with increasing maternal age and body mass index, increased with fetal CRL and maternal serum free β -hCG and PAPP-A MoM and was lower in women of South Asian racial origin than in Caucasians and in pregnancies conceived by assisted reproduction techniques than in natural conceptions.

The median fetal fraction in pregnancies with fetal trisomy 21 was not significantly different from that in unaffected pregnancies. In trisomies 18 and 13, the fetal fraction was significantly reduced and this decrease could be explained by the association of these trisomies with low serum PAPP-A and free β -hCG, reflecting the smaller placental source of fetal cfDNA in maternal blood.

In 3% of pregnancies the cfDNA test failed to provide a result after first sampling and the main reason for such failure was low fetal fraction of < 4%. The rate of a failed result was similar in unaffected and trisomy-21 cases, but was increased in trisomy 18 and 13 pregnancies. Logistic regression analysis demonstrated that the risk of a failed cfDNA test was inevitably affected by the same factors as those affecting fetal fraction. Thus, the rate of a failed result increased with increasing maternal age and body mass index, decreased with increasing maternal serum level of free β -hCG and PAPP-A MoM and was higher in women of South Asian racial origin than in Caucasians and in pregnancies conceived by assisted reproduction techniques than in natural conceptions.

The options for the management of pregnancies with a failed cfDNA test result after first sampling include repeat cfDNA testing, invasive testing or no further investigation; similarly, the options for women with a second failed cfDNA test are invasive testing or no further investigation. An important determinant for selecting the appropriate management option is the estimated risk for trisomies from the combined test and the presence of sonographic features suggestive of trisomies 18 and 13. In this study, only 7% (22/316) with a failed cfDNA test chose to have invasive testing and in 91% of these

cases the estimated risk for trisomies from the combined test was ≥ 1 in 100. In contrast, in the patients with a failed cfDNA test, after either first or repeat testing, who decided against invasive testing, only 13% had an estimated risk for trisomies from the combined test of ≥ 1 in 100.

Comparison of findings to those in previous studies

The inverse association between fetal fraction and maternal body mass index, which could be attributed to a dilutional effect, but also an increase in maternal cfDNA with increasing weight, is compatible with the results of previous cfDNA studies^{12–15}. Similarly, our finding of a linear association between fetal fraction and serum free β -hCG and PAPP-A MoM provides further support to our suggestion that, since all three are produced by the placenta, their maternal serum levels provide an indirect measure of placental mass^{12–14}. A three-dimensional ultrasound study reported that, in trisomy-21 pregnancies, placental volume at 11–13 weeks' gestation was not significantly different from that in euploid pregnancies, but in trisomies 18 and 13 placental volume was decreased¹⁶.

In previous studies we found that the fetal fraction is decreased in women of African racial origin, primarily because of an increase in maternal cfDNA level rather than a decrease in fetal cfDNA in maternal blood^{12–14}. In this larger study, we found in the univariate analysis that, in women of African racial origin, the fetal fraction was significantly lower than in Caucasians but in the multivariate analysis this significance was lost. In contrast, in women of South Asian racial origin, the fetal fraction was significantly reduced both in the univariate and multivariate analyses and this decrease may be a consequence of an increase in maternal cfDNA level, rather than a decrease in fetal cfDNA in maternal blood¹⁴.

The finding that, in pregnancies conceived by assisted reproduction techniques, the fetal fraction is lower than in natural conceptions is compatible with a previous report

in multiple pregnancies¹⁷ and this may be the consequence of the degree of impaired placentation which can also explain the higher incidence of associated pregnancy complications, such as pre-eclampsia¹⁸.

A few studies compared fetal fraction in pregnancies with fetal trisomies 21, 18 or 13 with that in euploid pregnancies. Rava *et al.* examined high-risk pregnancies undergoing invasive testing at 10–23 weeks' gestation, including 160 euploid pregnancies and 90, 38 and 16 with trisomies 21, 18 and 13, respectively¹⁹. The mean fetal fraction was significantly higher in trisomy 21 (13.5%) than in euploid pregnancies (12.6%) and was significantly lower in trisomies 18 (8.9%) and 13 (9.0%) than in euploid pregnancies (12.6%). Dar *et al.* reported the results of screening at a median gestational age of 13 (range, 9–41) weeks in 17 885 pregnancies, including 140 with trisomy 21, 27 with trisomy 18 and 8 with trisomy 13²⁰. The median fetal fraction was 10.1% and this increased with gestational age and decreased with maternal weight; after adjustment for these variables the median MoM fetal fraction in trisomy 21 (1.05 MoM) was significantly higher and in trisomies 18 (0.92 MoM) and 13 (0.76 MoM) was lower than in euploid pregnancies (1.0 MoM). Palomaki *et al.* reported the results of a case–control study at a median gestational age of 15 (range, 8–22) weeks in 2157 pregnancies, including 212 cases with trisomy 21, 62 with trisomy 18 and 12 with trisomy 13²¹. The median fetal fraction in trisomy 21 (15.5%) was higher and in trisomy 18 (9.4%) was lower than in euploid pregnancies (13.3%); the value in trisomy 13 was 13.6%. Kinnings *et al.* reported the results of screening at a median gestational age of 13 (range, 10–40) weeks in 140 377 pregnancies, including 2214 with trisomy 21, 835 with trisomy 18 and 432 with trisomy 13²². The median fetal fraction increased with gestational age, decreased with maternal weight and was affected by the fetal aneuploidy status; the median value was 9.6% for euploid and trisomy 21 pregnancies, 8.2% for trisomy 18 and 8.7% for trisomy 13. The study also demonstrated that the fetal fraction in trisomic, compared to euploid pregnancies, changed with gestational age; the fetal fraction was initially lower for all three trisomies but then became higher than in euploid pregnancies after 16, 21 and 18 weeks for trisomies 21, 18 and 13, respectively²².

Implications for clinical practice

On the basis of the results from this and previous studies, it can be concluded that, in trisomies 18 and 13, but not in trisomy 21, the fetal fraction is lower and the rate of failed cfDNA test is higher than in unaffected pregnancies. Consequently, pregnancies with a failed test can be considered as being at increased risk for trisomies 18 and 13, but not for trisomy 21. However, the results of the study of Kinnings *et al.* suggest that this problem of over-representation of trisomies 18 and 13 in pregnancies with a failed cfDNA test is confined to the first half of pregnancy²².

The management of pregnancies with a failed cfDNA test should depend essentially on the reason for carrying out such a test in the first place, as well as the cost of the cfDNA test; however, most companies will repeat the test at no additional cost. If there was prior screening with a low-risk result, the preferred option would be to repeat the cfDNA test and explain to the parents that such testing would provide a result in > 60% of cases. Some patients would prefer to avoid any further testing because of the associated anxiety; in these patients and in those with a failed second cfDNA test it would be advisable to carry out a detailed ultrasound scan for features of trisomies 18 and 13 and in the presence of such features invasive testing should be considered. If prior screening had provided a high-risk result but there are no ultrasound features of aneuploidy, most patients would prefer repeat cfDNA testing but a few would select to have invasive testing.

Conclusions

In cases of failed cfDNA test, fetal trisomies 18 and 13, but not trisomy 21, are over-represented. In the management of such cases, the decision in favor of invasive testing should depend on the risk from prior screening and the results of detailed ultrasound examination.

ACKNOWLEDGMENT

The study was supported by a grant from The Fetal Medicine Foundation (UK Charity No: 1037116).

Disclosure

The cost of collection and analysis of the samples for the cell-free DNA test in the NHS hospitals was covered by Ariosa Diagnostics, Inc. San Jose, CA, USA. This organization had no role in study design, data collection, data analysis, data interpretation or writing of the report.

REFERENCES

1. Gil MM, Quezada MS, Revello R, Akolekar R, Nicolaides KH. Analysis of cell-free DNA in maternal blood in screening for fetal aneuploidies: updated meta-analysis. *Ultrasound Obstet Gynecol* 2015; **45**: 249–266.
2. Pergament E, Cuckle H, Zimmermann B, Banjevic M, Sigurjonsson S, Ryan A, Hall MP, Dodd M, Lacroite P, Stosic M, Chopra N, Hunkapiller N, Prosen DE, McAdoo S, Demko Z, Siddiqui A, Hill M, Rabinowitz M. Single-nucleotide polymorphism-based noninvasive prenatal screening in a high-risk and low-risk cohort. *Obstet Gynecol* 2014; **124**: 210–218.
3. American College of Obstetricians and Gynecologists. Cell-free DNA screening for fetal aneuploidy. Committee Opinion No. 640. *Obstet Gynecol* 2015; **126**: e31–37.
4. Gil MM, Revello R, Poon LC, Akolekar R, Nicolaides KH. Clinical implementation of routine screening for fetal trisomies in the UK NHS: cell-free DNA test contingent on results from first-trimester combined test. *Ultrasound Obstet Gynecol* 2016; **47**: 45–52.
5. Quezada MS, Gil MM, Francisco C, Orós G, Nicolaides KH. Screening for trisomies 21, 18 and 13 cell-free DNA analysis of maternal blood at 10–11 weeks' gestation and the combined test at 11–13 weeks. *Ultrasound Obstet Gynecol* 2015; **45**: 36–41.
6. Robinson HP, Fleming JE. A critical evaluation of sonar crown rump length measurements. *Br J Obstet Gynaecol* 1975; **82**: 702–710.
7. Wright D, Kagan KO, Molina FS, Gazzoni A, Nicolaides KH. A mixture model of nuchal translucency thickness in screening for chromosomal defects. *Ultrasound Obstet Gynecol* 2008; **31**: 376–383.

8. Kagan KO, Wright D, Spencer K, Molina FS, Nicolaides KH. First-trimester screening for trisomy 21 by free beta-human chorionic gonadotropin and pregnancy-associated plasma protein-A: impact of maternal and pregnancy characteristics. *Ultrasound Obstet Gynecol* 2008; **31**: 493–502.
9. Sparks AB, Struble CA, Wang ET, Song K, Oliphant A. Noninvasive prenatal detection and selective analysis of cell-free DNA obtained from maternal blood: evaluation for trisomy 21 and trisomy 18. *Am J Obstet Gynecol* 2012; **206**: 319.e1–9.
10. Ashoor G, Syngelaki A, Wagner M, Birdir C, Nicolaides KH. Chromosome-selective sequencing of maternal plasma cell-free DNA for first-trimester detection of trisomy 21 and trisomy 18. *Am J Obstet Gynecol* 2012; **206**: 322.e1–5.
11. Ashoor G, Syngelaki A, Wang E, Struble C, Oliphant A, Song K, Nicolaides KH. Trisomy 13 detection in the first trimester of pregnancy using a chromosome-selective cell-free DNA analysis. *Ultrasound Obstet Gynecol* 2013; **41**: 21–25.
12. Ashoor G, Poon L, Syngelaki A, Mosimann B, Nicolaides KH. Fetal fraction in maternal plasma cell-free DNA at 11–13 weeks' gestation: Effect of maternal and fetal factors. *Fetal Diagn Ther* 2012; **31**: 237–243.
13. Ashoor G, Poon L, Syngelaki A, Mosimann B, Nicolaides KH. Fetal fraction in maternal plasma cell-free DNA at 11–13 weeks' gestation: relation to maternal and fetal characteristics. *Ultrasound Obstet Gynecol* 2013; **41**: 26–32.
14. Poon LC, Musci T, Song K, Syngelaki A, Nicolaides KH. Maternal plasma cell-free fetal and maternal DNA at 11–13 weeks' gestation: relation to fetal and maternal characteristics and pregnancy outcomes. *Fetal Diagn Ther* 2013; **33**: 215–223.
15. Palomaki GE, Kloza EM, Lambert-Messerlian GM, Haddow JE, Neveux LM, Ehrich M, van den Boom D, Bombard AT, Deciu C, Grody WW, Nelson SF, Canick JA. DNA sequencing of maternal plasma to detect Down syndrome: An international clinical validation study. *Genet Med* 2011; **13**: 913–920.
16. Wegrzyn P, Faro C, Falcon O, Peralta CF, Nicolaides KH. Placental volume measured by three-dimensional ultrasound at 11 to 13+ 6 weeks of gestation: relation to chromosomal defects. *Ultrasound Obstet Gynecol* 2005; **26**: 28–32.
17. Bevilacqua E, Gil MM, Nicolaides KH, Ordoñez E, Cirigliano V, Dierickx H, Willems PJ, Jani JC. Performance of screening for aneuploidies by cell-free DNA analysis of maternal blood in twin pregnancies. *Ultrasound Obstet Gynecol* 2015; **45**: 61–66.
18. Chaveeva P, Carbone IF, Syngelaki A, Akolekar R, Nicolaides KH. Contribution of method of conception on pregnancy outcome after the 11–13 weeks scan. *Fetal Diagn Ther* 2011; **30**: 9–22.
19. Rava RP, Srinivasan A, Sehnert AJ, Bianchi DW. Circulating fetal cell-free DNA fractions differ in autosomal aneuploidies and monosomy X. *Clin Chem* 2014; **60**: 243–250.
20. Dar P, Curnow KJ, Gross SJ, Hall MP, Stosic M, Demko Z, Zimmermann B, Hill M, Sigurjonsson S, Ryan A, Banjevic M, Kolacki PL, Koch SW, Strom CM, Rabinowitz M, Benn P. Clinical experience and follow-up with large scale single-nucleotide polymorphism-based noninvasive prenatal aneuploidy testing. *Am J Obstet Gynecol* 2014; **211**: 527.e1–17.
21. Palomaki GE, Kloza EM, Lambert-Messerlian GM, van den Boom D, Ehrich M, Deciu C, Bombard AT, Haddow JE. Circulating cell free DNA testing: are some test failures informative? *Prenat Diagn* 2015; **35**: 289–293.
22. Kinnings SL, Geis JA, Almasri E, Wang H, Guan X, McCullough RM, Bombard AT, Saldivar JS, Oeth P, Deciu C. Factors affecting levels of circulating cell-free fetal DNA in maternal plasma and their implications for noninvasive prenatal testing. *Prenat Diagn* 2015; **35**: 816–822.

RESUMEN

Objetivos Primero, informar sobre la distribución de la fracción de ADN fetal en sangre materna (cfDNA, por sus siglas en inglés) y la tasa del resultado de una prueba fallida de cfDNA en las trisomías 21, 18 y 13, en comparación con embarazos no afectados por estas trisomías; segundo, examinar los posibles efectos de las características maternas y fetales en la fracción fetal; y tercero, considerar las opciones para el manejo posterior de los embarazos con un resultado fallido.

Métodos Éste fue un estudio de cohorte de 10 698 embarazos únicos sometidos a un cribado para las trisomías fetales 21, 18 y 13 mediante la prueba de cfDNA a las 10–14 semanas de gestación. Se obtuvieron 160 casos con trisomía 21, 50 con trisomía 18, 16 con trisomía 13 y 10 472 no se vieron afectados por estas trisomías. Se utilizó un análisis de regresión multivariante para determinar los predictores significativos de la fracción fetal y del resultado fallido de la prueba de cfDNA a partir de características maternas y fetales.

Resultados La fracción fetal disminuyó con el aumento del índice de masa corporal y la edad materna, fue menor en las mujeres con etnicidad del sur de Asia que en las caucásicas y en las fecundaciones asistidas, en comparación con las espontáneas. La fracción fetal aumentó con la longitud céfalo-caudal (coronilla-rabadilla) del feto y con mayores niveles en el suero de la proteína plasmática A asociada al embarazo y la hormona gonadotrópica coriónica humana (subunidad B). La mediana de la fracción fetal fue del 11,0% (rango intercuartílico (RIC), 8,3–14,4%) en el grupo no afectado, del 10,7% (RIC, 7,8–14,3%) en la trisomía 21, del 8,6% (RIC, 5,0–10,2%) en la trisomía 18 y del 7,0% (RIC, 6,0–9,4%) en la trisomía 13. Se encontró un resultado fallido de la prueba de cfDNA después de la primera toma de muestras en el 2,9% del grupo no afectado, el 1,9% de la trisomía 21, el 8,0% de la trisomía 18 y el 6,3% de la trisomía 13. En los casos con resultado fallido, el 7% de las mujeres tuvieron pruebas invasivas, principalmente debido al riesgo elevado de la prueba combinada y/o la presencia de características ecográficas que sugerían trisomías 18 y 13. Todos los casos de trisomías se detectaron prenatalmente.

Conclusiones En los casos de una prueba de cfDNA fallida, la tasa de las trisomías 18 y 13, pero no la de la trisomía 21, es mayor que en aquellos con una prueba exitosa. En el manejo de estos casos, la decisión a favor de las pruebas invasivas debería depender del riesgo de cribado previo y los resultados de un examen ecográfico detallado.

目的: 首先, 报道与未发现 21, 18, 13 三体的孕妇相比, 21, 18, 13 三体孕妇中游离 (cell-free, cf) DNA 胎儿比值的分布以及 cfDNA 检测失败率; 其次, 研究孕妇和胎儿特点对胎儿比值的可能影响; 第三, 考虑对检测失败的孕妇的进一步处理方法。

方法: 本研究对 10 698 例单胎妊娠进行队列研究, 采用 cfDNA 检测, 对孕 10~14 周的孕妇进行胎儿 21, 18, 13 三体筛查。160 例为 21 三体, 50 例为 18 三体, 16 例为 13 三体, 10 472 例为阴性结果。采用多变量回归分析, 确定孕妇和胎儿特点中胎儿比值和 cfDNA 检测失败的显著预测因子。

结果: 胎儿比值随体重指数和孕妇年龄增加而降低, 南亚人与白种人相比胎儿比值较低, 人工受孕与自然受孕相比胎儿比值较低。胎儿比值随胎儿头臀长增加以及血清妊娠相关血浆蛋白 A 和游离 β -人绒毛膜促性腺激素水平升高而增大。胎儿比值中位数在阴性结果组为 11.0% [四分位数间距 (interquartile range, IQR), 8.3%~14.4%], 21 三体组为 10.7% (IQR, 7.8%~14.3%), 18 三体组为 8.6% (IQR, 5.0%~10.2%), 13 三体组为 7.0% (IQR, 6.0%~9.4%)。阴性结果组、21 三体组、18 三体组和 13 三体组中, 分别有 2.9%、1.9%、8.0%和 6.3%的孕妇在首次取样后 cfDNA 检测失败。在检测失败的病例中, 7%的孕妇进行了侵入性检查, 主要原因是联合筛查显示为高危孕妇和 (或) 超声检查提示 18 和 13 三体。所有三体病例均产前检测出。

结论: cfDNA 检测失败的病例中, 18 和 13 三体 (而非 21 三体), 所占的比率高于检测成功的病例。处理这类病例时, 是否进行侵入性检查取决于之前筛查确定的风险以及详细的超声检查结果。